DDR

Synergism Between Tramadol and Meloxicam in the Formalin Test Involves Both Opioidergic and Serotonergic Pathways

Mario A. Isiordia-Espinoza,¹ Flavio Terán-Rosales,¹ Gerardo Reyes-García,¹ and Vinicio Granados-Soto^{2*}

¹Sección de Estudios de Postgrado e Investigación, Escuela Superior de Medicina del Instituto Politécnico Nacional, México, D.F., 11340 México

²Departamento de Farmacobiología, Centro de Investigación y de Estudios Avanzados (Cinvestav), Sede Sur. México, D.F., 14330 México

Strategy, Management and Health Policy					
Enabling Technology, Genomics, Proteomics		Preclinical Development Toxicology, Formulation Drug Delivery, Pharmacokinetics		Postmarketing Phase IV	

ABSTRACT This study was designed to evaluate the antinociceptive interaction of the tramadol-meloxicam combination in different proportions (tramadol + meloxicam in 1:1, 1:3, and 3:1 ratios), as well as the role of nitric oxide, opioidergic, and serotonergic pathways in the antinociceptive effect of the combination. The effects of individual drugs and fixed-ratio combinations were assayed using the 3% formalin test in mice. Isobolographic analysis was employed to characterize the synergism produced by the combinations. Tramadol (3.16-10 mg/kg, i.m.), meloxicam (3.16-17.8 mg/kg, i.m.), and tramadolmeloxicam combinations produced a dose-dependent antinociceptive effect. ED₃₀ values were estimated for the individual drugs, and isobolograms were constructed. The tramadol + meloxicam 1:1 and 1:3 ratio combinations showed synergistic interactions while the 3:1 ratio produced additive effects. Naloxone (1 mg/kg, i.m.) or methiothepin (0.1 mg/kg, i.m.), but not L-NAME (3 mg/kg, i.m.), prevented the antinociceptive effects of the combination. These data suggest that (1) the tramadol-meloxicam combination produces a functional synergistic interaction that involves both opioid and serotonin receptors, and (2) this combination may be a promising tool in pain management. Drug Dev Res 73: 43–50, 2012. © 2011 Wiley Periodicals, Inc.

Key words: tramadol; meloxicam; synergism; opioid receptors; serotonin receptors

INTRODUCTION

Opioids remain the most effective therapy available for the treatment of moderate to severe pain in humans. However, the problems arising from unwanted side effects persist. Thus, combinations of opioids and other analgesic drugs are commonly used to control postoperative pain. The potential advantage of using combination therapy is that the analgesic effects can be maximized, whereas the incidence of side effects could be minimized. In addition, the multiplicity of mechanisms involved in pain suggests

Grant sponsor: Escuela Superior de Medicina del Instituto Politécnico Nacional; Grant number: 20100823.

*Correspondence to: Vinicio Granados-Soto, Departamento de Farmacobiología, Cinvestav, Sede Sur, Calzada Tenorios 235, Col. Granjas Coapa, D.F., 14330 México. E-mail: vgranados@prodigy.net.mx

Received 13 April 2011; Accepted 12 June 2011

Published online in Wiley Online Library (wileyonlinelibrary.com). DOI: 10.1002/ddr.20461

that combination therapy can improve pain management [Raffa, 2001].

Tramadol is a synthetic, centrally acting, analgesic agent widely used for pain relief in children and adults [Scott and Perry, 2000]. It is effective in moderate to severe postoperative pain with an overall efficacy similar to that of morphine or alfentanil. Previous clinical studies have shown that co-administration of magnesium, ketamine [Unlügenç et al., 2002], ketorolac [Pieri et al., 2002], or acetyl salicylate [Pang et al., 2000] and tramadol improves analgesia and patient comfort and decreases the amount of tramadol required for pain management. Animal studies supporting these interactions are lacking. Meloxicam is a non-steroidal antiinflammatory drug (NSAID) of the enolic acid class of oxicam derivatives indicated for the treatment of rheumatoid arthritis, osteoarthritis and other joint diseases [Engelhardt, 1996; Euller-Ziegler et al., 2001]. It acts mainly through inhibition of cyclooxygenase-2 [Laird et al., 1997; Pairet et al., 1998]. The present study was designed to assess the possible synergistic interaction between tramadol and meloxicam after intramuscular administration by isobolographic analyses. In addition, the possible role of nitrergic, opioidergic, and serotonergic pathways in the synergy induced by the tramadolmeloxicam combination was also assessed.

MATERIALS AND METHODS

Animals

Male Balb/c mice aged 8–9 weeks and weighing 20–25 g were used. The mice were housed at 22°C with a 12-h/12-h light/dark cycle. Animals had free access to food and tap water up to the time of the experiment. All experiments were conducted in accordance with the Guidelines on Ethical Standards for Investigation of Experimental Pain in Animals [Zimmerman, 1983]. In addition, the study was approved by our local Ethics Committee.

Drugs

Tramadol was obtained from Grünenthal de México, S.A. de C.V. (Mexico City, México), and meloxicam was a gift of Senosiain, S.A. de C.V. (Celaya, México). L-NG-nitroarginine methyl ester (L-NAME), naloxone, and methiothepin were obtained from Sigma (St. Louis, MO). All drugs were dissolved in sterile saline.

Measurement of Nociceptive Activity

Nociception was assessed using the formalin test. Mice were placed in clear plastic chambers with a mirror placed at a 45-degree angle to allow an unobstructed view of the paw. The injection was made into the plantar surface of the right hindpaw with $30 \,\mu$ l of dilute 3% formalin using a 30-gauge needle. Animals

were then returned to the chambers; nociceptive behavior was observed immediately after formalin injection. Nociceptive behavior was quantified as the licking time on the injected paw. Mice were sacrificed in a CO_2 chamber at the end of the experiment.

Experimental Design

Different groups were used to characterize the dose-response curve of the various drugs. Increasing doses of tramadol (3.16, 5.6, 7.5, and 10 mg/kg) or meloxicam (3.16, 5.6, 10, and 17.8 mg/kg) were given i.m. 20 min before s.c. administration of 3% formalin. Controls were administered saline solution. Once the dose-response curve of each drug was obtained, an experimental ED_{30} value was determined for each drug. The tramadol-meloxicam combination was evaluated in different proportions (tramadol + meloxicam in 1:1, 1:3, and 3:1 ratios). To assess the possible mechanism(s) of action for the combination, L-NAME (3 mg/kg), naloxone (1 mg/kg), methiothepin (0.1 mg/kg), or vehicle were administered i.p. 10 min before the tramadol+meloxicam combination (ED₃₀ value); 50 min later, formalin was injected.

Data Analysis

Data are presented as mean \pm SEM for ≥ 6 animals per group. The total time of licking corresponding to the second phase of the assay was determined from 15–45 min with regard to formalin administration. Dose-response data are presented as the percentage antinociception of the total licking time on the second phase of the formalin test. The percentage antinociception was calculated according to the following equation [Argüelles et al., 2002]:

[(Vehicle - postcompound)/vehicle] \times 100.

Dose-response curves were constructed and the experimental points fitted using least-squares linear regression. The SE estimate was calculated as described by Tallarida [2000].

Isobolographic analysis is a convenient tool for evaluating the interaction between analgesic drugs [Argüelles et al., 2002; Tallarida, 2000]. In the present study, we used this technique to determine the nature of interactions between tramadol and meloxicam. Isobolographic analysis assumes that the combination of drugs is made from equipotent doses of the individual drugs. Thus, from the dose-response curves of each individual agent, the dose resulting in 50% of the effect (ED₅₀ value) can be determined. However, considering that a maximal effect of 100% as the total suppression of formalin-induced licking and that meloxicam was unable to achieve a 50% response, the calculation of an ED₅₀ value was not feasible. Therefore, the ED_{30} value was estimated instead of the ED_{50} value. Subsequently, a dose-response curve was obtained by concurrent delivery of the two drugs in a constant dose ratio (fixed-ratio) based on the ED_{30} values of each individual agent. The ED_{30} value was evaluated for three combinations (tramadol+meloxicam in 1:1, 1:3, and 3:1 ratios). From the resulting dose-response curve of the combination, the experimental ED_{30} value was then calculated.

To determine whether the interaction between two drugs given in combination was synergistic, additive, or antagonistic, the theoretical additive ED_{50} value (Z_{add}) was estimated from the doseresponse curves of each drug administered individually, considering that the observed effect with the combination results of the sum of the individual effects of each component. This theoretical ED_{30} value was then compared with the experimental ED_{30} value (Z_{exp}) to determine whether there is a statistically significant difference [Tallarida et al., 1999; Tallarida, 2002].

The theoretical and experimental ED_{30} values of the studied combinations were also contrasted by calculating the interaction index (γ) as follows:

 $\gamma = ED_{30}$ value of combination (experimental)/ ED₃₀ value of combination (theoretical).

The interaction index indicates the portion of the ED_{30} value of individual drugs that accounts for the corresponding ED_{30} value in the combination. Values of ~ 1 correspond to an additive interaction, values of > 1 imply an antagonistic interaction, and values of < 1 indicate a synergistic interaction.

Statistical Analysis

Dose-response data were analyzed by one-way analysis of variance (ANOVA) followed by the Student–Newman–Keuls test for post hoc comparison. The theoretical additive ED_{30} and the experimentally derived ED_{30} values were evaluated using Student's *t*-test. An experimental ED_{30} value significantly lower than the theoretical additive ED_{30} value was considered to indicate a synergistic interaction between tramadol and meloxicam. Mechanisms of action (control group compared with the antagonist group) were evaluated by one-way ANOVA followed by the Student–Newman–Keuls test. Statistical significance was considered to be achieved when P < 0.05.

RESULTS

Antinociceptive Effects of Tramadol, Meloxicam, and Tramadol + Meloxicam Combinations

Tramadol and meloxicam significantly reduced formalin-induced licking in mice (Fig. 1). Figures 2A,B

shows dose-response curves for these drugs as well as their combinations during the second phase of the formalin test. The individual drugs and the combinations decreased the nociceptive behavior in a dosedependent manner, reaching a maximal effect of \sim 80.8%, 52.12%, and 57.23% for tramadol, meloxicam, and the tramadol + meloxicam 1:3 combination, respectively).

Isobolographic Analysis

The maximum effect reached by the greatest dose of the tramadol + meloxicam combinations in the 1:1, 3:1, and 1:3 ratios were approximately 53%, 46%, and 57%, respectively. Of note, the sum of the individual effects (ED₃₀ value of each drug) suggests that the tramadol doses would contribute 30% of its maximum effect (80.8%), i.e., 24.2%. Likewise, if it is assumed

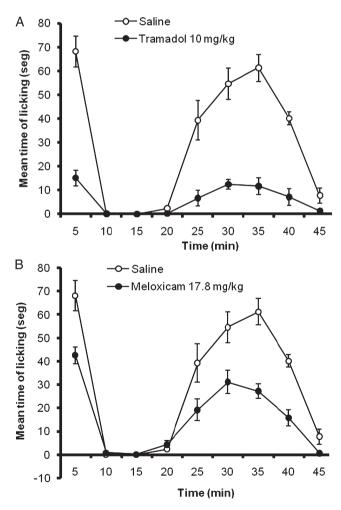
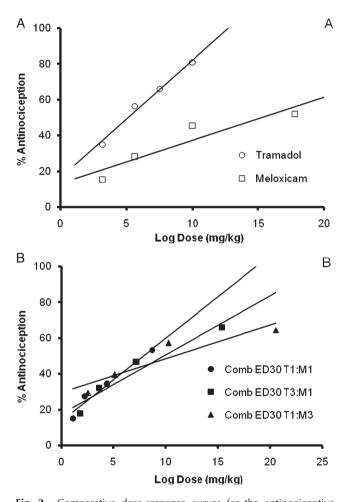


Fig. 1. Time course of the antinociceptive effect of tramamol (10 mg/kg, i.m., **A**) or meloxicam (17.8 mg/kg, i.m., **B**) in mice submitted to the 3% formalin test. Data are expressed as the mean time of licking \pm SEM of \geq 6 animals.

that the maximum effect of meloxicam was 52.1%, the ED_{30} value of this drug would contribute 15.63% of the effect of the combination according to the experimental test. The algebraic sum of such effects would be around 39.8%, which is less than the maximum effect of each combination. Thus, the tramadol + meloxicam combination in 1:1 and 1:3, but not 3:1, ratios produced the greatest effect (Fig. 3A,C). Accordingly, the experimental ED_{30} values of the tramadol + meloxicam combinations in 1:1 and 1:3 ratios were lower compared with the theoretical additive ED_{30} value of the combination (Fig. 3A and C, Table 1). Furthermore, analysis of the interaction index showed an



increase in potency for the tramadol+meloxicam combinations in 1:1 ($\gamma = 0.61$) and 1:3 ($\gamma = 0.55$), but not 3:1 ($\gamma = 0.89$) ratio.

Mechanism of Action

L-NAME was unable to reverse the antinociceptive effect of the combination (Fig. 4A). In contrast, naloxone and methiothepin significantly reduced the antinociceptive effect of the tramadol–meloxicam combination (Fig. 4B,C).

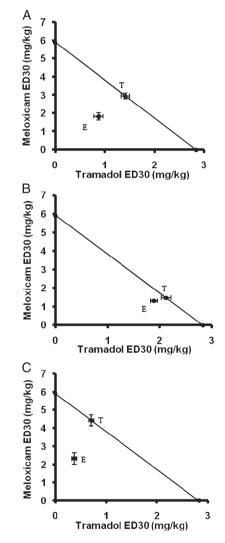


Fig. 2. Comparative dose-response curves for the antinociceptive effect of tramadol and meloxicam alone (**A**) or combined (**B**) during the second phase of the formalin test. Doses of tramadol (\bigcirc) were 3.1, 5.6, 7.5, and 10 mg/kg (i.m.), whereas those of meloxicam (\square) were 3.1, 5.6, 10 and 17.8 mg/kg. Doses of the tramadol+meloxicam combination in 1:1 ratio (\bullet) were 2.8, 1.4, 0.7, and 0.4+5.9, 2.9, 1.5, and 0.7 mg/kg, respectively. Doses of the tramadol+meloxicam combination in 3:1 ratio (\blacksquare) were 4.2, 2.1, 1.1 and 0.5+2.9, 1.5, 0.7, and 0.4 mg/kg, respectively. Doses of the tramadol+meloxicam combination in 1:3 ratio (\blacktriangle) were 1.4, 0.7, 0.4, and 0.2+8.8, 4.4, 2.2 and 1.1 mg/kg, respectively.

Fig. 3. Isobolograms showing the interaction between tramadol and meloxicam (1:1 **[A]**, 3:1 **[B]**, and 1:3 **[C]** ratio) in the mice formalin test. Horizontal and vertical bars indicate SEM. The oblique line between the *x* and *y* axes are the theoretical additive line. The point in the middle of this line, indicated by T, is the theoretical additive point calculated from the individual drug ED_{30} values. The point indicated by E is the actually observed ED_{30} value with the combination. In all cases, the experimental ED_{30} value point is situated below the additive line, being significantly different for the theoretical ED_{30} value, indicating a significant synergism (P < 0.05).

	T+M, 1:1 ratio ED ₃₀ values	T+M, 3:1 ratio ED ₃₀ values	T+M, 1:3 ratio ED ₃₀ values
Z _{add} (mg/kg)	4.36 ± 0.25	3.6 ± 0.16	5.13 ± 0.36
$Z_{\rm exp}$ (mg/kg)	$2.69 \pm 0.27^*$	3.2 ± 0.02	$2.85 \pm 0.05^*$
Interaction index	0.61	0.89	0.55

TABLE 1. Theoretical (Z_{add}) and Experimental (Z_{exp}) ED₃₀ Values ± SEM for the Tramadol (T)/Meloxicam (M) Combination in Different Proportions.

*P < 0.05 vs Z_{add}, by the Student's *t*-test.

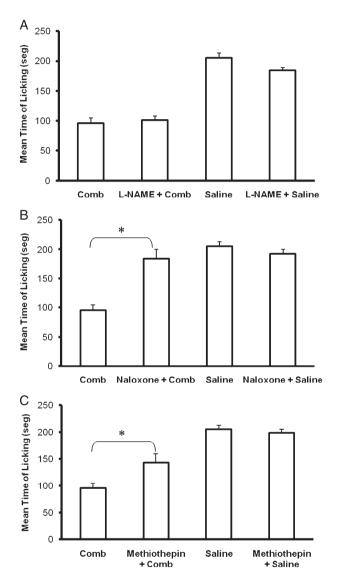


Fig. 4. Effect of L-NAME (**A**), naloxone (**B**), and methiothepin (**C**) on the antinociceptive effect of the tramadol–meloxicam combination (1:1 ratio). Bars are the mean \pm SEM for at least 6 animals. *Significantly different (*P*<0.05) from the combination (Comb), by one-way ANOVA followed the Student–Newman–Keuls test.

DISCUSSION

The current study demonstrates that tramadol produces dose-dependent antinociception in the formalin test. The antinociceptive effect of tramadol has been shown in several pain animal models, including the formalin test. Therefore, our results are in agreement with previous observations indicating that tramadol produces antinociception after systemic administration [Chen et al., 2002; Granados-Soto and Argüelles, 2005; Pozos-Guillen et al., 2006]. Systemic administration of meloxicam produced a dose-related antinociceptive effect during the second phase of the assay. Our results agree with previous studies showing that systemic meloxicam is able to reduce nociception in several pain animal models [Engelhardt et al., 1995; Laird et al., 1997; Santos et al., 1998; Pinardi et al., 2003; Dudhgaonkar et al., 2008]. The findings of the study also confirm that opioids and nonsteroidal antiinflammatory drugs (NSAIDs) show different profiles of antinociceptive activity, as tramadol exhibited greater antinociceptive potency and efficacy (Fig. 2A).

The present study focused on the nature of the interaction between tramadol and meloxicam in different proportions. Previous studies have shown that tramadol is able to increase the effect of adrenergic and serotonergic drugs [Pinardi et al., 1998], ketamine [Chen et al., 2002], metamizol [Poveda et al., 2003], naproxen [Satyanarayana et al., 2004], gabapentin [Granados-Soto and Argüelles, 2005], rofecoxib [García-Hernández et al., 2007], and desketoprofen [Miranda and Pinardi, 2009]. However, to our knowledge, this is the first report regarding synergistic interaction between tramadol and meloxicam. Our results confirm several observations showing that co-administration of opioids and NSAIDs leads to a synergistic interaction in inflammatory [Fletcher et al., 1997; Jiménez-Andrade et al., 2003; Poveda et al., 2003] and acute [Chen et al., 2002; Miranda et al., 2007] pain models in mice and rats as well as to an opioid-sparing effect in humans [Silvanto et al., 2002].

Particularly, in the present study, isobolographic analyses demonstrated a significant synergistic interaction between tramadol and meloxicam for the proportions 1:1, 1:3, but not for 3:1, which resulted in an additive effect. It is interesting to note that, according to the proportion of the drug in the combination, a synergistic interaction can became additive. This fact suggests that the proportion of drugs is an important feature for the synergistic effect of the combination as previously shown [Berenbaum, 1989; Chou, 2006; Miranda and Pinardi, 2009] and strongly supports the need to assess different drug ratios when evaluating drug interactions.

The synergism observed between tramadol and meloxicam supports the general premise of interactions between analgesic drugs that act through different mechanisms of action [Berenbaum, 1989; Chou, 2006]. Tramadol is a weak opioid that also inhibits norepinephrine and serotonin reuptake [Driessen et al., 1993; Bamigbade et al., 1997; Oliva et al., 2002], whereas meloxicam is a cyclooxygenase-2 preferring inhibitor [Laird et al., 1997; Pairet et al., 1998]. Moreover, there is evidence that other mechanisms of action participate in the antinociceptive effects of these drugs. For instance, µ-opioid receptor agonists inhibit activation of adenylyl cyclase [Ingram and Williams, 1996] and release of substance P and calcitonin gene-related peptide from primary afferent neurons [Yaksh, 1988]; they open K^+ channels leading to hyperpolarization, reduction in firing of the primary afferent neuron, and antinociception [Yoshimura and North, 1983; Rodrigues and Duarte, 2000]. Meloxicam activates the nitric oxide–cyclic GMP pathway [Aguirre-Bañuelos and Granados-Soto, 2000], Ca^{2+} -activated K⁺ channels [Ortiz et al., 2005], and the cholinergic inhibitory descendent system [Miranda et al., 2003] in the formalin test. The fact that naloxone (an opioid antagonist) and methiothepin (a 5-HT_{1/2/6/7} receptor antagonist [Hoyer et al., 1994]) reduce the antinociceptive effect of the combination strongly suggests that at least some of these mechanisms participate in the observed synergy with tramadol and meloxicam.

Nitric oxide, opioid, and serotonergic mechanisms were analyzed by testing the effects of L-NAME, naloxone, and methiothepin on tramadol/meloxicaminduced antinociception. The local antinociceptive effect of the combination was unaffected by the nitric oxide synthesis inhibitor L-NAME [Gibson et al., 1990], thus precluding the involvement of the nitric oxide pathway in the effect of the combination. The lack of effect could not be attributed to the dose of L-NAME used, as this dose has been shown to reduce tolerance to morphine-induced antinociception [Homayoun et al., 2003]. This result seems surprising, as the peripheral antinociceptive of meloxicam is diminished by L-NAME in rats submitted to the formalin test [Aguirre-Bañuelos and Granados-Soto, 2000]. This difference could be attributable to the administration route. In contrast, systemic naloxone diminished the antinociceptive activity of the tramadolmeloxicam combination. Our data are in line with several observations indicating that tramadol, but not meloxicam, activates μ opioid as well as α_2 adrenoceptors [Raffa et al., 1992; Kayser et al., 1992; Ide et al., 2006]. In addition, systemic administration of methiothepin significantly reduced combination-induced antinociception. These data suggest that tramadol [Bamigbade et al., 1997; Oliva et al., 2002] and meloxicam, as is the case for other NSAIDs [Björkman, 1995; Pini et al., 1995, 1996], may interact with the spinal serotonergic system by inhibiting the reuptake or increasing release of spinal 5-HT. 5-HT could target specific 5-HT receptors in the spinal cord. Since methiothepin is a high-affinity 5-HT_{1/2/6/7} receptor antagonist [Hoyer et al., 1994], the present data suggest that these receptors could be involved in combination-induced antinociception in the formalin test. More specifically, the candidate spinal receptor could be either 5-HT_{1/2} receptors, linked to spinal antinociception [Oyama et al., 1996; Sasaki et al., 2001], but not 5-HT_{6/7} receptors, as their spinal activation is associated with pronociception [Rocha-González et al., 2005; Castañeda-Corral et al., 2009]. However, on the basis of this experiment, the possible participation of other types of spinal 5-HT receptors cannot be ruled out. Together, these data suggest that the tramadol-meloxicam combination activates opioid and serotonergic receptors to produce antinociception in the formalin test.

In conclusion, the present study demonstrated that tramadol and meloxicam produce antinociception in the formalin test after i.m. administration. Moreover, the data indicate the presence of a functional synergistic interaction between tramadol and meloxicam that involves the opioid and serotonergic system.

ACKNOWLEDGMENTS

The authors greatly appreciate the technical and bibliographic assistance of Guadalupe C. Vidal-Cantú and Héctor Vázquez, respectively. Mario A. Isiordia-Espinoza is a Conacyt fellow. This work is part of the doctoral dissertation of Mario A. Isiordia-Espinoza.

REFERENCES

- Aguirre-Bañuelos P, Granados-Soto V. 2000. Evidence for the participation of the nitric-oxide-cyclic GMP pathway in the antinociceptive action of meloxicam in the formalin test. Eur J Pharmacol 395:9–13.
- Argüelles CF, Torres-López JE, Granados-Soto V. 2002. Peripheral antinociceptive action of morphine and the synergistic interaction with lamotrigine. Anesthesiology 96:921–925.
- Bamigbade TA, Davidson C, Langford RM, Stamford JA. 1997. Actions of tramadol, its enantiomers and principal metabolite, O-desmethyltramadol, on serotonin (5-HT) efflux and uptake in the rat dorsal raphe nucleus. Br J Anaesthesia 79:352–356.
- Berenbaum MC. 1989. What is synergy? Pharmacol Rev 41:93-141.

- Björkman R. 1995. Central antinociceptive effects of non-steroidal anti-inflammatory drugs and paracetamol. Experimental studies in the rat. Acta Anaesthesiol Scand 103(Suppl):1–44.
- Castañeda-Corral G, Rocha-González HI, Araiza-Saldaña CI, Ambriz-Tututi M, Vidal-Cantú GC, Granados-Soto V. 2009. Role of peripheral and spinal 5-HT₆ receptors according to the rat formalin test. Neuroscience 162:444–452.
- Chen Y, Chan SY, Ho PC. 2002. Isobolographic analysis of the analgesic interactions between ketamine and tramadol. J Pharm Pharmacol 54:623–631.
- Chou TC. 2006. Theoretical basis, experimental design, and computerized simulation of synergism and antagonism in drug combination studies. Pharmacol Rev 58:621–681.
- Driessen B, Reimann W, Giertz H. 1993. Effects of the central analgesic tramadol on the uptake and release of noradrenaline and dopamine in vitro. Br J Pharmacol 108:806–811.
- Dudhgaonkar SP, Tandan SK, Kumar D, Arunadevi R, Prakash VR. 2008. Synergistic interaction between meloxicam and aminoguanidine in formalin-induced nociception in mice. Eur J Pain 12: 321–328.
- Engelhardt G. 1996. Pharmacology of meloxicam, a new nonsteroidal anti-inflammatory drug with an improved safety profile through preferential inhibition of COX-2. Br J Rheumatol 35(Suppl 1):4–12.
- Engelhardt G, Homma D, Schlegel K, Utzmann R, Schnitzler C. 1995. Anti-inflammatory, analgesic, antipyretic and related properties of meloxicam, a new non-steroidal anti-inflammatory agent with favourable gastrointestinal tolerance. Inflamm Res 44: 423–433.
- Euller-Ziegler L, Vélicitat P, Bluhmki E, Türck D, Scheuerer S, Combe B. 2001. Meloxicam: a review of its pharmacokinetics, efficacy and tolerability following intramuscular administration. Inflamm Res 50(Suppl 1):S5–S9.
- Fletcher D, Benoist JM, Gautron M, Guilbaud G. 1997. Isobolographic analysis of interactions between intravenous morphine, propacetamol, and diclofenac in carrageenin-injected rats. Anesthesiology 87:317–326.
- García-Hernández L, Déciga-Campos M, Guevara-López U, López-Muñoz FJ. 2007. Co-administration of rofecoxib and tramadol results in additive or sub-additive interaction during arthritic nociception in rat. Pharmacol Biochem Behav 87: 331–340.
- Gibson A, Mirzazadeh S, Hobbs AJ, Moore PK. 1990. L-NGmonomethyl arginine and L-NG-nitro arginine inhibit nonadrenergic, non-cholinergic relaxation of the mouse anococcygeus muscle. Br J Pharmacol 99:602–606.
- Granados-Soto V, Argüelles CF. 2005. Synergic antinociceptive interaction between tramadol and gabapentin after local, spinal and systemic administration. Pharmacology 74:200–208.
- Homayoun H, Khavandgar S, Mehr SE, Namiranian K, Dehpour AR. 2003. The effects of FK506 on the development and expression of morphine tolerance and dependence in mice. Behav Pharmacol 14: 121–127.
- Hoyer D, Clarke DE, Fozard JR, Hartig PR, Martin GR, Mylecharane EJ, Saxena PR, Humphrey PP. 1994. International union of pharmacology classification of receptors for 5-hydroxytryptamine (serotonin). Pharmacol Rev 46:157–203.
- Ide S, Minami M, Ishihara K, Uhl GR, Sora I, Ikeda K. 2006. Mu opioid receptor-dependent and independent components in effects of tramadol. Neuropharmacology 51:651–658.

- Ingram SL, Williams JT. 1996. Modulation of the hyperpolarizationactivated current (Ih) by cyclic nucleotides in guinea-pig primary afferent neurons. J Physiol 492:97–106.
- Jiménez-Andrade JM, Ortiz MI, Pérez-Urizar J, Aguirre-Bañuelos P, Granados-Soto V, Castañeda-Hernández G. 2003. Synergistic effects between codeine and diclofenac after local, spinal and systemic administration. Pharmacol Biochem Behav 76:463–471.
- Kayser V, Besson JM, Guilbaud G. 1992. Evidence for a noradrenergic component in the antinociceptive effect of the analgesic agent tramadol in an animal model of clinical pain, the arthritic rat. Eur J Pharmacol 224:83–88.
- Laird JM, Herrero JF, García de la Rubia P, Cervero F. 1997. Analgesic activity of the novel COX-2 preferring NSAID, meloxicam in mono-arthritic rats: central and peripheral components. Inflamm Res 46:203–210.
- Miranda HF, Puig MM, Dursteler C, Prieto JC, Pinardi G. 2007. Dexketoprofen-induced antinociception in animal models of acute pain: synergy with morphine and paracetamol. Neuropharmacology 52:291–296.
- Miranda HF, Lemus I, Pinardi G. 2003. Effect of the inhibition of serotonin biosynthesis on the antinociception induced by nonsteroidal anti-inflammatory drugs. Brain Res Bull 61: 417–425.
- Miranda HF, Pinardi G. 2009. Lack of effect of naltrexone on the spinal synergism between morphine and non steroidal antiinflammatory drugs. Pharmacol Reports 61:268–274.
- Oliva P, Aurilio C, Massimo F, Grella A, Maione S, Grella E, Scafuro M, Rossi F, Berrino L. 2002. The antinociceptive effect of tramadol in the formalin test is mediated by the serotonergic component. Eur J Pharmacol 445:179–185.
- Ortiz MI, Castañeda-Hernández G, Granados-Soto V. 2005. Pharmacological evidence for the activation of Ca^{2+} -activated K^+ channels by meloxicam in the formalin test. Pharmacol Biochem Behav 81:725–731.
- Oyama T, Ueda M, Kuraishi Y, Akaike A, Satoh M. 1996. Dual effect of serotonin on formalin-induced nociception in the rat spinal cord. Neurosci Res 25:129–135.
- Pairet M, van Ryn J, Schierok H, Mauz A, Trummlitz G, Engelhardt G. 1998. Differential inhibition of cyclooxygenases-1 and -2 by meloxicam and its 4'-isomer. Inflamm Res 47:270–276.
- Pang W, Huang S, Tung CC, Huang MH. 2000. Patient-controlled analgesia with tramadol versus tramadol plus lysine acetyl salicylate. Anesth Analg 91:1226–1229.
- Pieri M, Meacci L, Santini L, Santini G, Dollorenzo R, Sansevero A. 2002. Control of acute pain after major abdominal surgery in 585 patients given tramadol and ketorolac by intravenous infusion. Drugs Exp Clin Res 28:113–118.
- Pinardi G, Pelissier T, Miranda HF. 1998. Interactions in the antinociceptive effect of tramadol in mice: an isobolographic analysis. Eur J Pain 2:343–350.
- Pinardi G, Sierralta F, Miranda HF. 2003. Atropine reverses the antinociception of nonsteroidal anti-inflammatory drugs in the tail-flick test of mice. Pharmacol Biochem Behav 74:603–608.
- Pini LA, Sandrini M, Vitale G. 1995. Involvement of brain serotonergic system in the antinociceptive action of acetylsalicylic acid in the rat. Inflamm Res 44:30–35.
- Pini LA, Sandrini M, Vitale G. 1996. The antinociceptive action of paracetamol is associated with changes in the serotonergic system in the rat brain. Eur J Pharmacol 308:31–40.

- Poveda R, Planas E, Pol O, Romero A, Sánchez S, Puig MM. 2003. Interaction between metamizol and tramadol in a model of acute visceral pain in rats. Eur J Pain 7:439–448.
- Pozos-Guillén JA, Aguirre-Bañuelos P, Arellano-Guerrero A, Castañeda-Hernández G, Hoyo-Vadillo C, Pérez-Urizar J. 2006. Isobolographic analysis of the dual-site synergism in the antinociceptive response of tramadol in the formalin test in rats. Life Sci 79:2275–2282.
- Raffa RB, Friderichs E, Reimann W, Shank RP, Codd EE, Vaught JL. 1992. Opioid and nonopioid components independently contribute to the mechanism of action of tramadol, an "atypical" opioid analgesic. J Pharmacol Exp Ther 260:275–285.
- Raffa RB. 2001. Pharmacology of oral combination analgesics: rational therapy for pain. J Clin Pharm Ther 26:257–264.
- Rocha-González HI, Meneses A, Carlton SM, Granados-Soto V. 2005. Pronociceptive role of peripheral and spinal 5-HT₇ receptors in the formalin test. Pain 117:182–192.
- Rodrigues AR, Duarte ID. 2000. The peripheral antinociceptive effect induced by morphine is associated with ATP-sensitive K^+ channels. Br J Pharmacol 129:110–114.
- Santos AR, Vedana EM, De Freitas GA. 1998. Antinociceptive effect of meloxicam, in neurogenic and inflammatory nociceptive models in mice. Inflamm Res 47:302–307.
- Sasaki M, Ishizaki K, Obata H, Goto F. 2001. Effects of 5-HT_2 and 5-HT_3 receptors on the modulation of nociceptive transmission in rat spinal cord according to the formalin test. Eur J Pharmacol 424:45–52.
- Satyanarayana PS, Jain NK, Singh A, Kulkarni SK. 2004. Isobolographic analysis of interaction between cyclooxygenase

inhibitors and tramadol in acetic acid-induced writhing in mice. Prog Neuropsychopharmacol Biol Psychiatry 28: 641–649.

- Scott LJ, Perry CM. 2000. Tramadol: a review of its use in perioperative pain. Drugs 60:139–176.
- Silvanto M, Lappi M, Rosenberg PH. 2002. Comparison of the opioid-sparing efficacy of diclofenac and ketoprofen for 3 days after knee arthroplasty. Acta Anaesthesiol Scand 46: 322–328.
- Tallarida RJ. 2000. Drug synergism and dose-effect data analysis. New York: Chapman & Hall/CRC. p 1–72.
- Tallarida RJ. 2002. The interaction index: a measure of drug synergism. Pain 98:163–168.
- Tallarida RJ, Stone DJ, McCarty JD, Raffa RB. 1999. Response surface analysis of synergism between morphine and clonidine. J Pharmacol Exp Ther 289:8–13.
- Unlügenç H, Gündüz M, Ozalevli M, Akman H. 2002. A comparative study on the analgesic effect of tramadol, tramadol plus magnesium, and tramadol plus ketamine for postoperative pain management after major abdominal surgery. Acta Anaesthesiol Scand 46:1025–1030.
- Yaksh TL. 1988. Substance P release from knee joint afferent terminals: modulation by opioids. Brain Res 458:319–324.
- Yoshimura M, North RA. 1983. Substancia gelatinosa neurons hyperpolarized in vitro by enkephalin. Nature 305:529–530.
- Zimmerman M. 1983. Ethical guidelines for investigations of experimental pain in conscious animals. Pain 16:109–110.